

# Mycoplasma Detection Kit

## Cell Culture Control

### Contamination & Controls

Cat.-No.	Amount
PP-401S	10 reactions
PP-401L	50 reactions

For general laboratory use

Quality guaranteed for 12 months

Store at -20°C

Aliquoting of reagents and handling on ice is recommended

### Description

Mycoplasma Detection Kit provides a highly sensitive, easy-to-perform and rapid tool for detection of Mycoplasma contaminations in cell cultures or other biological materials. The kit is based on the amplification of a conserved 16S rRNA coding region of *Mycoplasma* by PCR resulting in a characteristic 268 bp fragment. It allows the detection of all common *Mycoplasma* and *Ureaplasma* species with extreme sensitivity. Due to this sensitivity, please pay special attention to avoid cross contaminations.

### Table of tested species

Species	Origin
<i>Mycoplasma bovis</i>	Bovine
<i>Mycoplasma columborale</i>	Avian
<i>Mycoplasma bovigenitalium</i>	Bovine
<i>Mycoplasma iners</i>	Avian
<i>Mycoplasma gallinarum</i>	Avian
<i>Mycoplasma faecium</i>	Human
<i>Mycoplasma gallinaceum</i>	Mammalian/Avian
<i>Mycoplasma hominis</i>	Human
<i>Mycoplasma hyorhinis</i>	Porcine
<i>Mycoplasma synoviae</i>	Avian
<i>Ureaplasma urealyticum</i>	Human

### Kit contents

#### Hot Start Polymerase (red cap)

S pack: 7 µl

L pack: 30 µl

#### Master Mix (green cap)

Primer, dNTP's, Reaction Buffer

S pack: 250 µl

L pack: 1,25 ml

#### Control DNA (white cap)

S pack: 7 µl

L pack: 30 µl

#### Sample Buffer (blue cap)

S pack: 600 µl

L pack: 3 ml

### Additionally required material

- pipettes and filter tips
- PCR tubes
- micro centrifuge
- PCR thermal cycler
- agarose gel and electrophoresis system

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#### Protocol

##### Preparation of cell culture supernatant

Transfer 0.5 to 1 ml supernatant immediately prior splitting of the cells to a sterile vial. Growing the cells without antibiotics is not necessary.

- centrifuge samples for 30 sec at 250g
- transfer supernatant in a new vial and discard cell debris
- centrifuge for 15 min at 13.000-15.000g to sediment the mycoplasma
- decant carefully and discard supernatant
- resuspend the pellet (please note that the pellet may not be always visible) in 50 µl Sample Buffer and vortex well
- incubate the samples for 5 min at 95°C
- centrifuge the samples briefly and place them on ice

##### Preparation of other biological material

Testing of mycoplasma contaminations in sera, cryo cultures or cells requires the extraction of DNA prior to PCR. The use of a genomic DNA Extraction Kit is recommended.

#### PCR Reaktion

Prepare a Premix of the following components:

Premix	1 sample	5 samples
Master Mix	23,5 µl	117,5 µl
Polymerase	0,5 µl	2,5 µl

For each assay pipet 24 µl Premix in a PCR vial and add 1 µl of the prepared sample. For preparation of the positive control add 1 µl of Control DNA, as negative control apply 1 µl Sample Buffer. Mix and centrifuge the vials briefly. Place the vials in a thermocycler.

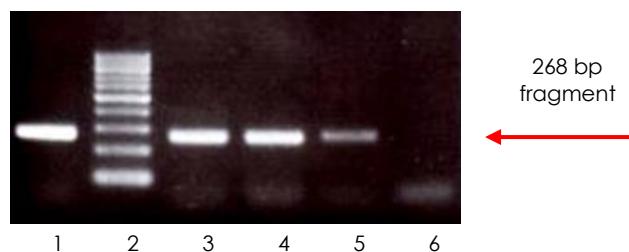
#### PCR program

Temperature	Time	Number of Cycles
94°C	2 min	35
94°C	30 sec	
55°C	30 sec	
72°C	30 sec	
72°C	2 min	1

#### Analysis of amplified products

- add 5 µl gel loading buffer to each vial, centrifuge and mix briefly
- load 5 µl of each assay onto a 2% agarose gel and run gel electrophoresis

#### Gel Analysis



- 1: positive control
- 2: 100 bp DNA Ladder
- 3 and 4: strongly contaminated samples
- 5: weakly contaminated sample
- 6: negative control

A gel band at approx. 270 bp is the indicator for a mycoplasma contamination of the sample.

#### References

- <http://de.wikipedia.org/wiki/Mycoplasmataceae>
- <http://de.wikipedia.org/wiki/Mykoplasmen>
- Uphoff, Drexler (2002) Comparative PCR analysis for detection of Mycoplasma infections in continuous cell lines. In Vitro Cell Dev. Biol. Anim. **38**: 79-85
- van Kuppeveld et al. (1992) Genus-Species-Specific Identification of Mycoplasmas by 16S rRNA Amplification. Applied and Environmental Microbiology, **58**: 2606-2615
- Winner, Rosengarten, Citti (2000) In vitro cell Invasion of Mycoplasma gallisepticum. Infect. Immun. **68**: 4238-4244
- Methodensammlung der Bund und Länder-Arbeitsgemeinschaft Gentechnik 7/2006