

## DATA SHEET



## Klentaq Pol Hot Start

Heat-activatable DNA polymerase for high specificity, chemically modified

*Thermus aquaticus*, recombinant, *E.coli*

Cat. Nº.	Amount
<input type="checkbox"/> POL-128XS	100 units
<input type="checkbox"/> POL-128S	250 units
<input type="checkbox"/> POL-128M	500 units
<input type="checkbox"/> POL-128L	1.000 units
<input type="checkbox"/> POL-128XL	2 x 1.000 units

### Unit Definition:

One unit is defined as the amount of the enzyme required to catalyze the incorporation of 10 nmoles of dNTPs into an acid-insoluble form in 30 minutes at 74 °C.

### Concentration:

5 units/ $\mu$ L

### Shipping:

Shipped on blue ice

### Storage Conditions:

Store at -20 °C

**For *in vitro* use only!**

### Additional Storage Conditions:

Avoid freeze/thaw cycles

### Shelf Life:

12 months

### Kit contents:

#### Klentaq Polymerase Hot Start (blue cap)

5 units/ $\mu$ L Klentaq Pol Hot Start in Tris-HCl pH 9.0 (25 °C), KCl, EDTA, 50% (v/v) Glycerol and stabilizers.

#### Klentaq Pol HS Reaction Buffer complete (red cap) - 10x conc.

Tris-HCl pH 8.0 (25°C) and stabilizers.

### Description:

Klentaq Pol Hot Start provides improved specificity and sensitivity when amplifying low-copy-number targets in complex backgrounds or when prolonged room-temperature set up is required. This ultra-pure enzyme, in addition to its hot-start capabilities, reduces false positives, amplifies a wide range of DNA sequence contexts. Klentaq Pol is purified by an additional separation process to reduce contaminating bacterial DNA sequences from the enzyme preparation. The polymerase activity is chemically blocked at ambient temperature and switched on automatically at the onset of the initial denaturation. The thermal activation prevents the extension of nonspecifically annealed primers and primer-dimer formation at low temperatures during PCR setup. The enzyme catalyzes the polymerization of nucleoside into duplex DNA in 5' 3' direction in the presence of magnesium. Klentaq lacks the 5' 3' exonuclease activity.

### Activation step:

Klentaq Hot Start Pol requires a prolonged heating or denaturing step. The chemical modification of the polymerase is reversed by the increased temperature of the hot start cycle.

### PCR Reaction Setup

The PCR procedure below shows appropriate volumes for a single 50  $\mu$ L reaction. For multiple reactions, prepare a master mix of components common to all and then dispense appropriate volumes into each PCR reaction tube prior to adding template DNA and primers.

Thaw, mix, and briefly centrifuge each component before use.

Add the following components to a microcentrifuge tube:

#### 1. Prepare PCR master mix

Note: Consider the volumes for all components listed next steps to determine the correct amount of water required to reach your final reaction volume.

Components	50 $\mu$ L rxn	[ final ]
Water, grade PCR	To 50 $\mu$ L	
10x Reaction Buffer	5 $\mu$ L	1X
dNTP (Mix 10 mM)	1 $\mu$ L	200 $\mu$ M
Klentaq Pol HS (5 U/ $\mu$ L)	0,5 $\mu$ L	2,5 U/reaction

Mix and briefly centrifuge the components.

#### 2. Add template DNA and primers

Components	50 $\mu$ L rxn	[ final ]
Forward primer (10 $\mu$ M)	0,5 - 2,5 $\mu$ L	0,1 - 0,5 $\mu$ M
Reverse primer (10 $\mu$ M)	0,5 - 2,5 $\mu$ L	0,1 - 0,5 $\mu$ M
DNA template		10 pg - 1 $\mu$ g**

\*\*genomic DNA: 1 ng-1 $\mu$ g; plasmidial or viral DNA: 1 pg-1 ng

Cap each tube, mix, and briefly centrifuge the content.

#### 3. Incubate reactions in a thermal cycler.

##### Recommended cycling conditions:

Step	Temp.	Time
Initial denaturation	95 °C	10 min
25 - 40 cycles	Denaturation	95 °C
	Annealing <sup>1</sup>	45-68 °C
	Elongation <sup>2</sup>	68 °C
Final extension (optional)	68 °C	1 - 2 min
Hold	4 - 8 °C	

1)The annealing temperature depends on the melting temperature of the primers used.

2)The elongation time depends on the length of the fragments to be amplified. A time of 2 min/kb is recommended.

For optimal specificity and amplification an individual optimization of the recommended parameters may be necessary for each new template DNA and/or primer pair.