



## 3'-End Oligonucleotide Labeling Reagent Kit

Cat. No.	Amount
APP-003	25 reactions x 50 µl (5 pmol each)

**For general laboratory use.**

**Shipping:** shipped on gel packs

**Storage Conditions:** store at -20 °C

**Additional Storage Conditions:** avoid freeze/thaw cycles

**Shelf Life:** 12 months

### Description:

3'-End Oligonucleotide Labeling Reagent Kit contains all buffer reagents required for efficient 3'-End Labeling of DNA oligonucleotides (length: 20 -100 bp, 5 pmol per reaction) except of oligonucleotide template to be labeled and labeled nucleotides..

The labeling principle is based on Terminal deoxynucleotidyl Transferase (TdT) that template-independently transfers labeled nucleotides to the 3'-OH group of ssDNA (e.g. an oligonucleotide) in the presence of  $\text{CoCl}_2$ . The number of nucleotide and thus label incorporation depends on the type of nucleotide (UTP/ddUTP) and type of label.

Labeled UTP: 1 – 3 label (average)

Labeled dUTP: multiple label (tail length is highly nucleotide specific)

Labeled ddUTPs: 1 label

The resulting 3'-End labeled oligonucleotides are ideally suited for applications involving sequence-specific protein binding or hybridization such as EMSA, Northern or Southern blots. Compared to internal, random labeled probes, the label is located at the 3'-End only and less likely interferes with probe binding.

TdT possesses a preference for single-stranded DNA (ssDNA) over dsDNA with 3'-overhangs or blunt ends. For the preparation of labeled dsDNA complexes, label each complementary oligonucleotide separately and anneal them before use.

### Content:

#### Terminal Deoxynucleotidyl Transferase (TdT)

30 µl (20 U/µl) in 100 mM potassium acetate (pH 6.8), 2 mM 2-mercaptoethanol, 0.01% Triton X-100 (v/v) and 50% glycerol (v/v)

#### 5x TdT Reaction Buffer

400 µl containing 1 M potassium cacodylate, 0.125 M Tris, 0.05% Triton X-100 (v/v), 5 mM  $\text{CoCl}_2$ , pH 7.2

#### Unlabeled Control Oligonucleotide (60 bp)

250 µl, 1 µM in 1x TE Buffer, pH 7.6

#### PCR-grade $\text{H}_2\text{O}$

12.5 ml

#### 1x TE Buffer, pH 7.6

100 ml containing 10 mM Tris-HCl, 1 mM EDTA, pH 7.6

#### Stop Buffer

400 µl, 0.5 M EDTA solution, pH 8



## 3'-End Oligonucleotide Labeling Reagent Kit

### 1. 3' End Oligonucleotide labeling reaction

- Store all components except of TdT on ice until use.
- Store TdT at -20°C until use.
- Final Assay volume: 50 µl
- Template requirements: oligonucleotide/ssDNA purified by HPLC or gel electrophoresis, 20 – 100 bp
- Add all components on ice exactly in the order listed below.
- Mix reaction gently by pipetting up and down. **Do not vortex!**
- Incubate 30 min at 37 °C.
- Add 1 µl Stop Buffer (0.5 M EDTA solution, pH 8) to stop each reaction.
- Store reactions on ice for subsequent use (see 3.) or -20 °C for long-term storage.

Component	Volume	Final concentration	Final molar amount
PCR grade H <sub>2</sub> O	31.5 µl	n/a	n/a
5x TdT Reaction Buffer	10 µl	1x	n/a
oligo-nucleotide template (1 µM)	5 µl	100 nM	5 pmol
Labeled UTP or ddUTP(10 µM)	2.5 µl	0.5 µM	50 pmol
TdT (20 U/µl)	1 µl	0.4 U/µl	20 U
Total volume	50 µl		

### 2. Estimation of labeling degree

Quantification of labeling degree is essential for reproducible downstream results.

Biotin or Digoxigenin-labeled oligonucleotides can be indirectly detected via Streptavidin or anti-Digoxigenin conjugates, respectively.

The labeling degree of fluorescent oligonucleotides can be directly detected by measurement of the nucleic acid-dye conjugate absorbance followed by a calculation of dye to base ratio according to the law of Lambert-Beer.